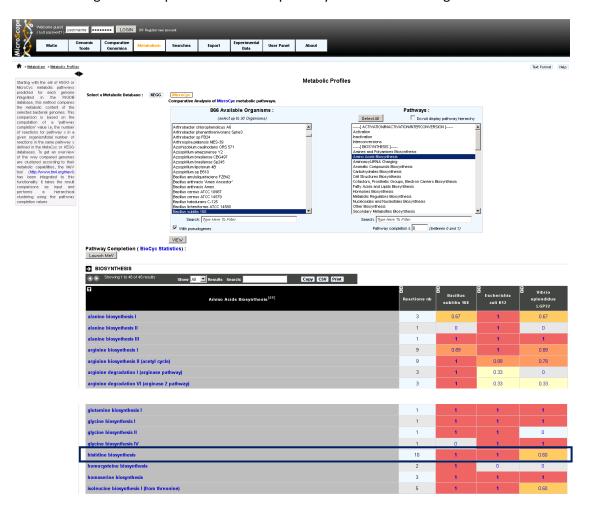
# Curation of metabolic data using the MicroScope platform: Corrected exercises

#### Ex. T-1: Metabolic profiles and co-evolved genes

Use the MicroCyc Metabolic profile tool for *E. coli* K12, *Bacillus subtilis* 168 and *Vibrio splendidus* LGP32. How many pathway holes are present in histidine biosynthesis pathway in LGP32? In the Metabolism section of Microscope main bar, select Metabolic profiles. In the Metabolic profiles page, select the genomes of interest and the pathways you want to compare. In the context of this exercise, we select Amino Acids biosynthesis, which includes histidine biosynthesis pathway. By clicking on the VIEW button, the results are displayed in the form of the degree of completion of selected pathways in the selected organisms:

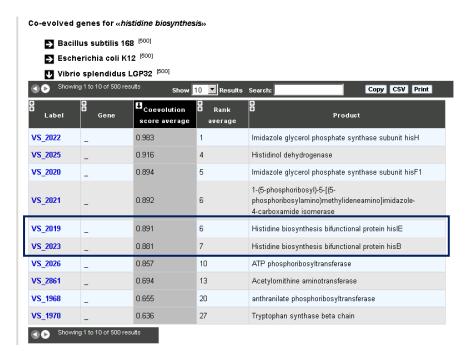


We can see that the pathway is incomplete in LGP32 (completion score < 1.00). By clicking on the histidine biosynthesis hyperlink, we can display the pathway table with the reactions and

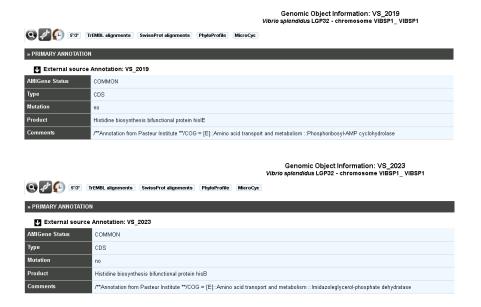
predicted genes in the selected genomes. We can see that **four enzymatic activities appear absents in LGT32:** 

	Reaction	s in « <i>histidin</i> e	e biosynthe	sis »	
Showing 1 to 10 of 10 results	Show 10 Results Search:	Copy CSV	Print		
O	Reactions	EC Number(s)	Bacillus Subtilis 168	Escherichia coli K12	Vibrio splendidus LGP32
ATPPHOSPHORIBOSYLTRANS-RXN	: ATP phosphoribosyltransferase	2.4.2.17	BSU34920 BSU34930	ECK2014	VS_2026
GLUTAMIDOTRANS-RXN: 2.4.2GL	UTAMIDOTRANS-RXN	2.4.2	BSU34870 BSU34890	ECK2018 ECK2020	VS_2020 VS_2022
HISTALDEHYD-RXN: Histidinal dehyd	Irogenase	1.1.1.23	BSU34910	ECK2015	VS_2025
HISTAMINOTRANS-RXN: histidinol-ph	nosphate transaminase	2.6.1.9	BSU22620	ECK2016	VS_1152 VS_2024
HISTCYCLOHYD-RXN: phosphoribos	yl-AMP cyclohydrolase	3.5.4.19	BSU34860	ECK2021	_
HISTIDPHOS-RXN: histidinol-phospha	atase	3.1.3.15	BSU29620	ECK2017	_
HISTOLDEHYD-RXN: Histidinol dehyd	drogenase	1.1.1.23	BSU34910	ECK2015	VS_2025
HISTPRATPHYD-RXN: phosphoribosy	rI-ATP diphosphatase	3.6.1.31	BSU34860	ECK2021	_
IMIDPHOSDEHYD-RXN: imidazolegly	cerol-phosphate dehydratase	4.2.1.19	BSU34900	ECK2017	_
PRIBFAICARPISOM-RXN: 1-(5-phosp 4-carboxamide isomerase	horibosyl)-5-(5-phosphoribosylamino)methylideneaminoimidazole	5.3.1.16	BSU34880	ECK2019	VS_2021
"histidine biosynthesis" MicroCyc Cross-spec "histidine biosynthesis" MetaCyc pathway	cies comparison				
Co-evolved genes for « <i>histidine</i>	biosynthesis»				
Bacillus subtilis 168 [500]					
<b>→</b> Escherichia coli K12 <sup>[500]</sup>					
→ Vibrio splendidus LGP32	[600]				
		CLOSE			

Using the Co-evolved genes in LGP32, find the candidate genes for missing enzymatic activities? Check blast results and synteny conservation to confirm your predictions: In the pathway table, we can display list of Co-evolved genes for histidine biosynthesis pathway in *V. splendidus* LGP32 in order to see if there are genes not initially associated to histidine biosynthesis but that shows significant co-evolution scores with other predicted genes of the pathway, indicative of a common evolutionary patterns of presence-absence across Microscope genomes that supports a common functional role:



There are two CDS (VS\_2019 and VS\_2023) that are not initially associated to the projected pathway but that shows the fifth and sixth best co-evolution scores among *V. splendidus* LGP32 CDSs with the other genes of the pathway (Notice that 1<sup>st</sup> to 5<sup>th</sup> best co-evolution scores corresponds with CDS already associated to histidine biosynthesis pathway). These CDS are annotated as Histidine biosynthesis bifunctional proteins not linked to any EC number in Microscope annotations, and as consequence are not initially associated to any step of Histidine biosynthesis pathway:



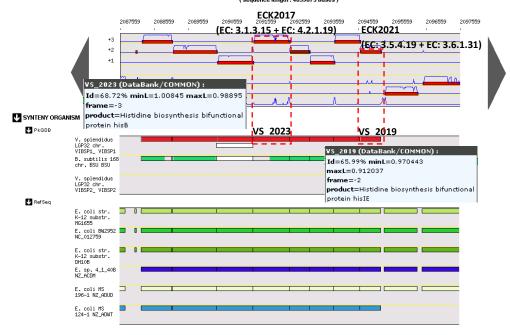
By comparing with the projections in *E. coli* K12, where the pathway is completely projected, we can see that missing enzymatic reactions in *V. splendidus* LGP32 corresponds to bifunctional enzymes in *E. coli* K12:

- **ECK2021 (HisIE):** Fused phosphoribosyl-AMP cyclohydrolase; phosphoribosyl-ATP pyrophosphatase (EC:3.5.4.19 + EC:3.6.1.31)
- **ECK2017 (HisB):** Fused histidinol-phosphatase; imidazoleglycerol-phosphate dehydratase (EC: 3.1.3.15 + EC: 4.2.1.19)

The analysis of genomic context conservation between *E. coli* K12 and *V. splendidus* LGP32 by selecting *E. coli* K12 as pivot genome in Microscope genome browser and *V. splendidus* LGP32 and *B. subtilis* 168 in the synteny maps confirms the predictions of co-evolution scores. **The** histidine biosynthesis operon of *E. coli* K12 is completely conserved in *V. splendidus* LGP32, with VS\_2019 being the counterpart of ECK2021 (Bifunctional EC: 3.5.4.19 + EC: 3.6.1.31) and VS\_2023 being the counterpart of ECK2017 (Bifunctional EC: 3.1.3.15 + EC: 4.2.1.19).

#### Escherichia coli K12 - chromosome U00096 2087559 -- 2097559

( sequence length : 4639675 bases )



<u>Ex. T-2</u>: Check if CanOE predicts a metabolon covering histidine biosynthesis pathway in *V. splendidus* LGP32: Following the same procedure as in Ex. T-1 to compare the metabolic profile of Histidine biosynthesis pathway in V. splendidus LGP32 and E. coli K12 and display the pathway table to compare GPR assignments in both genomes:

Reactions in « histidine biosynthesis » Copy CSV Print D Bacillus Vibrio coli K12 168 LGP32 BSU34920 ATPPHOSPHORIBOSYLTRANS-RXN: ATP phosphoribosyltransferase 2.4.2.17 ECK2014 VS 2026 BSU34870 ECK2018 VS\_2020 GLUTAMIDOTRANS-RXN: 2.4.2.- GLUTAMIDOTRANS-RXN 2.4.2.-HISTALDEHYD-RXN: Histidinal dehydrogenase 1.1.1.23 BSU34910 ECK2015 VS 2025 VS\_1152 HISTAMINOTRANS-RXN: histidinol-phosphate transaminase 2.6.1.9 VS 2024 HISTCYCLOHYD-RXN: phosphoribosyl-AMP cyclohydrolase 3.5.4.19 BSU34860 ECK2021 HISTIDPHOS-RXN: histidinol-phosphatase 3.1.3.15 BSU29620 ECK2017 HISTOLDEHYD-RXN: Histidinol dehydrogenase 1.1.1.23 BSU34910 ECK2015 VS 2025 HISTPRATPHYD-RXN: phosphoribosyl-ATP diphosphatase 3.6.1.31 BSU34860 ECK2021 IMIDPHOSDEHYD-RXN: imidazoleglycerol-phosphate dehydratase 4.2.1.19 BSU34900 ECK2017 PRIBFAICARPISOM-RXN: 1-(5-phosphoribosyl)-5-(5-phosphoribosylamino) methylidene aminoimidazole-proportional proportion of the proportionBSU34880 4-carboxamide isomerase Co-evolved genes for «histidine biosynthesis» Bacillus subtilis 168 [500] Escherichia coli K12 [500] > Vibrio splendidus LGP32 [500] CLOSE

On order to find if CanOE predicts any metabolon in V. splendidus LGP32 covering histidine biosynthesis pathway, go to Metabolism  $\rightarrow$  CanOE section of MicroScope main bar and search CanOE results by filtering on EC: 3.1.3.15:

CanOE
Fishing Candidate genes for Orphan Enzymes

Results Overa	II Organisms:				
<ul> <li>Consult global</li> </ul>	orphan reactions F	amily Level 💌			
Organism Spe	cific Results:	Escherichia coli K12		•	
C Consult metab	olon list for selected	l organism			
Consult orpha	n reactions for <b>selec</b>	ted organism Gene Le	vel 💌		
- 1					
Go					

Result of "search":

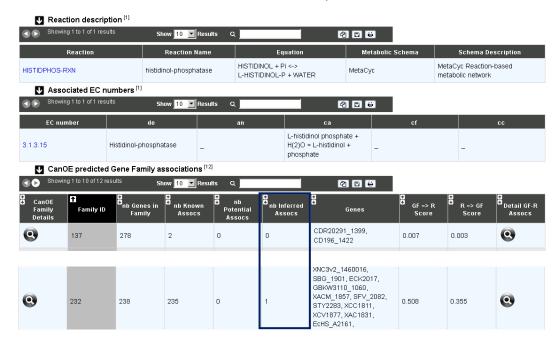
CanOE Search



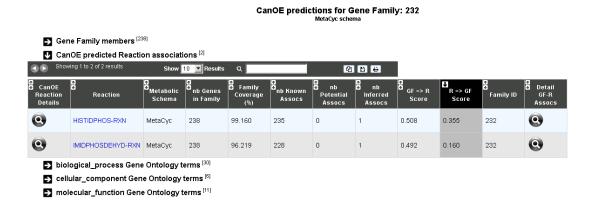
Click on the icon glass 'Canoe Reaction Details" and open "CanOE predicted gene associations":

We find this reaction associated to 12 different families defined by the CanOE method. Within these 12 families, there is only one of them (family ID 232; 238 genes, 235 "known" associations) for which CanOE strategy predicts a single inferred association, meaning that this association is included within a predicted metabolon (conserved genomic and metabolic contexts):

## CanOE predictions for Reaction: HISTIDPHOS-RXN MetaCyc schema



By clicking on the hyperlink of "CanOE family details" we can access the different reactions predicted for genes of this family included in metabolons. In this case, genes of this family are mainly associated to two different reactions that correspond to EC: 3.1.3.15 (HISTIDPHOS-RXN) and EC: 4.2.1.19 (IMIDPHOSDEHYD-RXN)

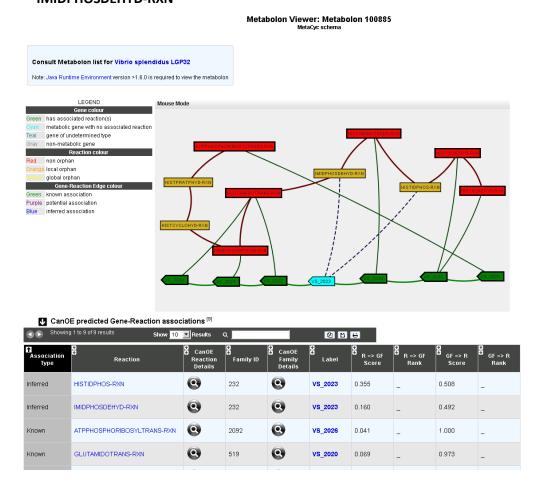


By clicking on the hyperlink "Detail GF-R assocs" for both reactions, we see that both reactions has been inferred by CanOE strategy associated to the same CDSs, which corresponds to VS\_2023 from V. splendidus LGP32 (single Inferred association by CanOE strategy associated to Gene Family 232):

### CanOE associations between: IMIDPHOSDEHYD-RXN / Family 232



□ In both cases, by clicking on "View metabolon" icon, we access to the predicted metabolon in V. splendidus LGP32, that covers histidine biosynthesis operon including the single inferred association of VS\_2023 with both HISTIDPHOS-RXN and IMIDPHOSDEHYD-RXN



So using the CanOE result we are able to find a gene candidate for the histidinol-phosphatase (EC 3.1.3.15) and the imidazoleglycerol-phosphate dehydratase (4.2.1.19) => VS\_2023 (Bifunctional enzyme as in E. coli).

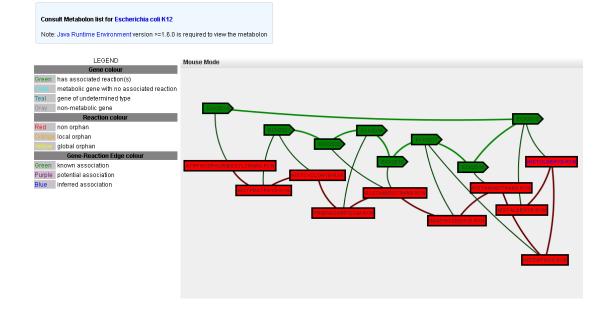
In comparison with the results of Ex. T-1, VS\_2019 is not included in the metabolon despite by analyzing genomic and metabolic context we found that corresponds to bifunctional phosphoribosyl-AMP cyclohydrolase; phosphoribosyl-ATP pyrophosphatase (EC:3.5.4.19 + EC:3.6.1.31). This can be explained by the fact that this gene is located in the boundaries of the metabolon, which covers from VS\_2020 to VS\_2026. In this context, CanOE requires that the metabolon should be flanked by known gene-reaction associations.

In comparison, in E. coli K12, where all gene-reaction associations are known and the genes are co-localized in an operonic structure, the predicted metabolon covers the whole histidine biosynthesis pathway:

Reactions in « histidine biosyn	thesis »	
Showing 1 to 10 of 10 results Show 10 Results Search:		Copy CSV Print
Reactions	O EC Number(s)	O Escherichia coli K12
ATPPHOSPHORIBOSYLTRANS-RXN: ATP phosphoribosyltransferase	2.4.2.17	ECK2014
GLUTAMIDOTRANS-RXN: 2.4.2GLUTAMIDOTRANS-RXN	24.2-	ECK2020 ECK2018
HISTALDEHYD-RXN: Histidinal dehydrogenase	1.1.1.23	ECK2015
HISTAMINOTRANS-RWI: histidinol-phosphate transaminase	2.6.1.9	ECK2016
HISTCYCLOHYD-RXN: phosphoribosyl-AMP cyclohydrolase	3.5.4.19	ECK2021
HISTIDPHOS-RXN: histidinol-phosphatase	3.1.3.15	ECK2017
HISTOLDEHYD-RXN: Histidinol dehydrogenase	1.1.1.23	ECK2015
HISTPRATPHYD-RXN: phosphoribosyl-ATP diphosphatase	3.6.1.31	ECK2021
IMIDPHOSDEHYD-RXIX: imidazoleglycerol-phosphate dehydratase	4.2.1.19	ECK2017
PRIBFAICARPISOM-R/Ni: 1-(5-phosphoribosyl)-5-(5- phosphoribosylamino)methylideneaminoimidazole-4-carboxamide isomerase	5.3.1.16	ECK2019
"hisbdine biosynthesid" MicroCyc pathway		

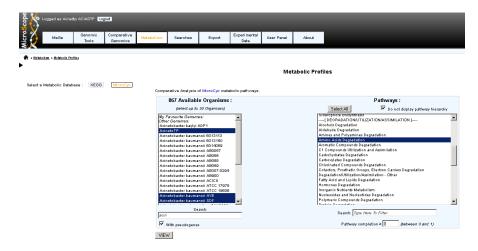
Metabolon Viewer: Metabolon 7477

MetaCvc schema

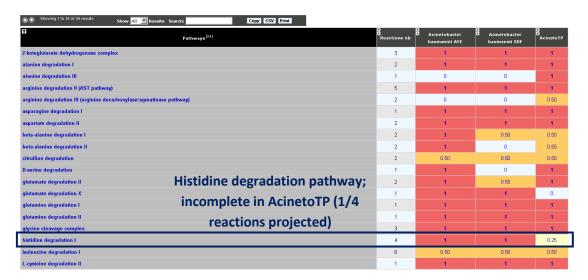


# Ex. A-1: Expert annotation of histidine degradation I pathway: Compare the metabolic profile of this pathway in AcinetoTP with A. baumannii AYE and A. baumannii SDF

In the Metabolism section of Microscope main bar, select Metabolic profiles. In the Metabolic profiles page, select the genomes of interest and the pathways you want to compare. In the context of this exercise, we select Amino Acids Degradation, which includes histidine degradation pathway



By clicking on the VIEW button, is displayed a table with rows representing pathways, columns representing genomes, and cells representing the completion index of the pathway in a given genome calculated as the ratio between number of projected reactions and total number of reactions in the pathway:

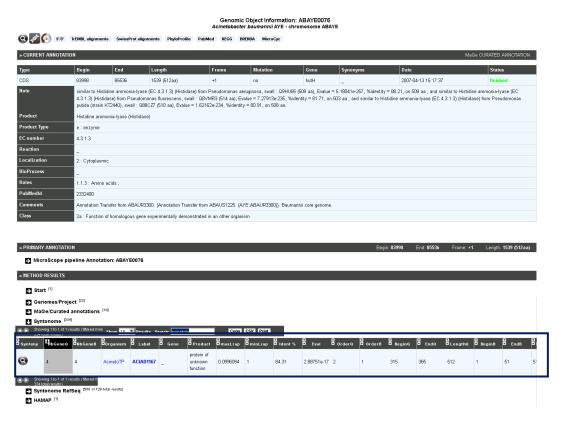


By clicking in the histidine degradation I hyperlink, a new window is displayed with the details of the pathway reactions and the corresponding CDS in the analysed genomes. We can see that only ACIAD0574 has been predicted in AcinetoTP associated to EC: 4.3.1.3 reaction (histidine ammonia-lyase). In contrast, the pathway is complete in *A. baumannii* AYE and *A. baumannii* SDF, with predicted genes co-linear in the same chromosome region:

		 	i ddalioi i i »		
Showing 1 to 4 of 4 results	Show 10 💌 Results Search:	Сору	CSV Print		
fi .	Reactions	EC Number(s)	DAcinetobacter baumannii AYE	DAcinetobacter baumannii SDF	AcinetoTP
FORMIMINOGLUTAMASE-RXN: formi	midoylglutamase	3.5.3.8	ABAYE0079	ABSDF3580	_
HISTIDINE-AMMONIA-LYASE-RXN: hi	stidine ammonia-lyase	4.3.1.3	ABAYE0076	ABSDF3583	ACIAD0574
IMIDAZOLONEPROPIONASE-RXN: ir	nidazolonepropionase	3.5.2.7	ABAYE0078	ABSDF3581	_
UROCANATE-HYDRATASE-RXN: uro	anate hydratase	4.2.1.49	ABAYE0075	ABSDF3584	_
"histidine degradation I' MicroCyc Cross-spec	ies comparison				

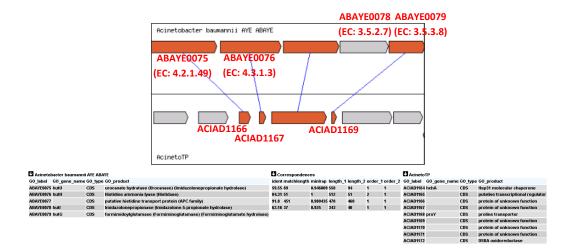
Reactions in « histidine degradation I »

Look at the conserved synteny between AcinetoTP and AYE. Is the histidine degradation operon conserved in AcinetoTP? In the pathway table, click on ABAYE0076 hyperlink to access to Microscope gene annotation interface. Go to the section *Syntonome* to explore the genome context conservation between A. baumannii AYE and AcinetoTP. Filter the Syntonome table by searching for AcinetoTP:

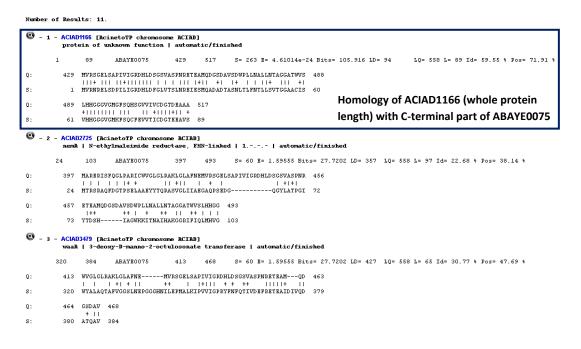


There is a synton of 4 orthologous genes between both genomes with a conserved genomic organization. By clicking on the glass icon, a graphical representation of the synton is displayed, where we can see that the histidine degradation operon in AcinetoTP is conserved but apparently highly degraded, with AcinetoTP genes that are shorter than *A. baumannii* AYE counterparts, consequence of a process of pseudogenization (gene inactivation):

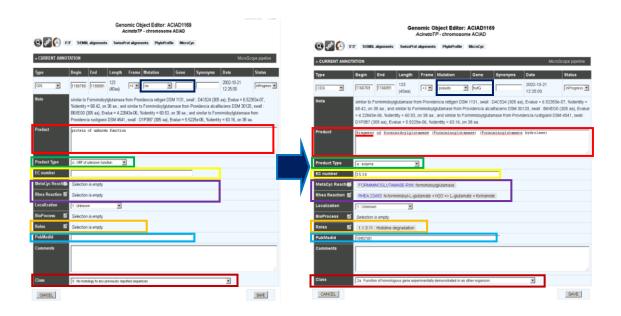
#### Synton #231\_1321\_6850\_6851



BLASTP analysis of A. baumannii AYE against AcinetoTP proteome confirms the pseudogenization of AcinetoTP genes involved in histidine degradation pathway. For example, BLASTP of ABAYE0075 (EC: 4.2.1.49; 558 amino acids length) against AcinetoTP proteome:



Based on these evidences, we can update annotation of ACIAD1166, ACIAD1167 and ACIAD1169 as pseudogenes of the corresponding orthologs in A. baumannii AYE, transferring the corresponding annotations from A. baumannii AYE to AcinetoTP CDSs and validating the corresponding MetaCyc and RHEA reactions, which can be retrieved by EC number synchronization. For example, for ACIAD1169, which corresponds to the pseudogenized form of ABAYE0079 (formimidoylglutamase; EC: 3.5.3.8):



Based on the results obtained, is this pathway functional in AcinetoTP? Validate the pathway accordingly by using the Pathway validation interface of Microscope: According to the previous results, we can conclude that histidine degradation pathway is non-functional in AcinetoTP consequence of a process of pseudogenization. As consequence, a "non-functional" status can be assigned in the Pathway curation interface of Microscope:



Ex. A-2: Expert annotation of glucarate/galactarate degradation pathway: Compare the metabolic profile of glucarate/galactarate degradation pathways in AcinetoTP with *E. coli* K12

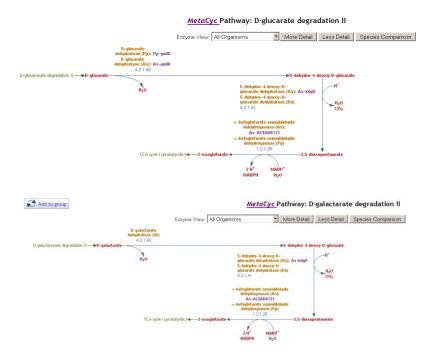
In the Metabolism section of Microscope main bar, select Metabolic profiles. In the Metabolic profiles page, select the genomes of interest and the pathways you want to compare. In the context of this exercise, we select Secondary Metabolites Degradation, which includes the

pathways for degradation of naturally occurring dicarboxylates D-glucarate and D-galactarate. We can see that there are two possible pathway variants for the degradation of each compound:

Ť	Secondary Metabolites Degradation <sup>[21]</sup>	Reactions nb	AcinetoTP	Escherichia coli K12
1,6-anhydr	o-N-acetylmuramic acid recycling	2	0.50	1
5-dehydro-	4-deoxy-D-glucuronate degradation	4	0.25	0.75
curcumin d	legradation	2	0	1
D-galactara	ite degradation l	4	0.50	1
D-galactara	ite degradation II	3	0.67	0.33
D-galacton	ate degradation	3	0	1
D-galacture	onate degradation I	4	0	1
D-glucarate	e degradation I	4	0.50	1
D-glucarate	e degradation II	3	0.67	0.33
fructoselys	ine and psicoselysine degradation	3	0	1
galactitol d	egradation	3	0	1
glucose de	gradation (oxidative)	5	0.40	0.20
ketoglucor	ate metabolism	8	0.63	0.75
L-galacton:	ate degradation	1	0	1
L-idonate o	legradation	3	0.67	1
mannitol de	egradation I	1	0	1

Refine the annotation of the CDSs ACIAD0126 to 0131 by using MicroScope tools and the publication PMID: 18364348: In this reference, the pathway of D-glucarate biosynthesis is experimentally verified in *A. baylyi* ADP1, demonstrating that it proceeds in a different manner as in *E. coli* K12. Whereas *E. coli* K12 degrades D-glucarate to 3-phosphoglycerate in 4 enzymatic steps (corresponds to D-glucarate degradation I pathway variant), *A. baylyi* ADP1 degrades D-glucarate to 2-oxoglucarate in 3 enzymatic steps, which corresponds to D-glucarate degradation II pathway variant:

In the same reference, knockout experiments in A. baylyi ADP1 identifies a cluster of genes comprising from ACIAD0126 to ACIAD0131 that are responsible of the transport and degradation of D-glucarate and D-galactarate, that is carried out by specific dehydratases for D-glucarate (EC:4.2.1.40) and D-galactarate (EC:4.2.1.42) that produces the common intermediate 5-dehydro-4-deoxy-D-glucarate, which is subsequently converted in 2-oxoglucarate in two reaction steps catalyzed by EC:4.2.1.41 and EC:1.2.1.26. This metabolic profile is represented in MetaCyc in two different pathway variants that correspond to D-glucarate degradation II and D-galactarate degradation II:

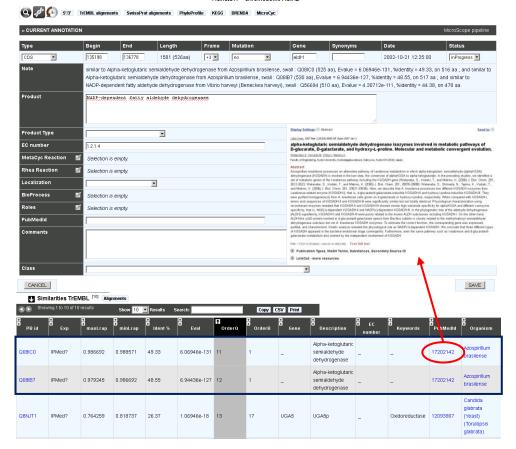


By comparing the metabolic profile of AcinetoTP for both pathway variants, in both cases there is 1 pathway hole that corresponds to EC: 1.2.1.26 activity:

Showing 1 to 3 of 3 results	Show 10 Results Search:		Сору	CSV Print
	Reactions	EC Number(s)	AcinetoTP	Escherichia coli K12
I.2.1.41-RXN: 5-dehydro-4-deoxyg	glucarate dehydratase	4.2.1.41	ACIAD0130	_
5-DIOXOVALERATE-DEHYDROG	ENASE-RXN: 2,5-dioxovalerate dehydrogenase	1.2.1.26	_	-
GALACTARDEHYDRA-RXN: galact	tarate dehydratase	4.2.1.42	ACIAD0126	ECK3116
-galactarate degradation (f* MicroCyc C -galactarate degradation (f* MetaCyc p:	cross-species comparison athway <b>Reactions in «</b> <i>D-glucarate degrac</i>	lation II »		COM DOWN
-galactarate degradation // MicroCyc C -galactarate degradation // MetaCyc pi Showing 1 to 3 of 3 results	Cross-species comparison athway		Сору	CSV Print
-galactarate degradation (f* MicroCyc C -galactarate degradation (f* MetaCyc p:	cross-species comparison athway <b>Reactions in «</b> <i>D-glucarate degrac</i>	ation II »  EC  Number(s)		CSV Print  CSV Print  CSV Escherichia  Coli K12
-galactarate degradation // MicroCyc C -galactarate degradation // MetaCyc pi Showing 1 to 3 of 3 results	cross-species comparison athway  Reactions in « <i>D-glucarate degrac</i> Show 10 Results Search:  Reactions	ec Ec	0	C O Escherichia
-galactarate degradation if MicroCyc C -galactarate degradation if MetaCyc pi	cross-species comparison athway  Reactions in « <i>D-glucarate degrac</i> Show 10 Results Search:  Reactions	G EC Number(s)	AcinetoTP	C O Escherichia

In the same reference is experimentally verified that 2-ketoglutarate semialdehyde dehydrogenase enzyme corresponding to EC: 1.2.1.26 is encoded by the gene ACIAD0131. This gene is annotated in MicroScope as NADP-dependent fatty aldehyde dehydrogenase associated to EC: 1.2.1.4. However, by analyzing the similarity profiles in BLASTP searches against SwissProt/TrEMBL entries with experimental evidence of their annotation, we find that the best hits in TrEMBL searches are against two *Azospirillum brasilense* entries annotated as Alpha-ketoglutaric semialdehyde dehydrogenase, with experimental evidence of this activity, although without EC number association.

# Genomic Object Editor: ACIAD0131 AcinetoTP - chromosome ACIAD



Based on these evidences, we should update the annotation of ACIAD0131 to Alpha-ketoglutaric semialdehyde dehydrogenase associated to EC: 1.2.1.26. The corresponding reactions in MetaCyc and RHEA should be also validated and the PubMed ID of the publication where the activity in A. baylyi ADP1 has been experimentally verified (PMID: 18364348):



Based on the results obtained, validate the 4 projected pathways accordingly by using the Pathway validation interface of Microscope: Accordingly with the evidences reported, we should assign a "Deleted" status to D-glucarate degradation I and D-galactarate degradation I

pathway variants (*E. coli* K12 variants not functional in AcinetoTP) in MicroScope Pathway Curation interface. In a similar manner, *D-glucarate degradation II* and *D-galactarate degradation II* should be assigned a "Validated" status (functional pathways in AcinetoTP).